

Sialyl Lewis X-Polysaccharide Conjugates: Targeting Inflammatory Lesions

Masahiro Sakagami, ^{1a}) Kazutoshi Horie, ^{1a}) Kazutaka Nakamoto, ^{1b})
Takayuki Kawaguchi, * ^{1c}) and Hiroshi Hamana * ^{1a})

Drug Delivery System Institute, Ltd., Noda-shi, Chiba 278, Japan

Received 21 July 1998; accepted 26 August 1998

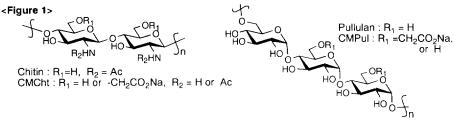
Abstract: A novel system for active targeting of inflammatory lesions has been established. A SLe^x-CMPul conjugate (2) showed accumulation that was 2.5-fold higher in inflammatory lesions *in vivo* than a SLN-CMPul conjugate (4) and 300-fold higher than monovalent SLe^x (6).

© 1998 Elsevier Science Ltd. All rights reserved.

In the last decade, the widespread occurrence of oligosaccharides in nature and their importance in various biological processes are becoming increasingly evident,²) but carbohydrate recognition has not been applied to targeting delivery to tissues or lesions, except for the interaction between galactose and the asialoglycoprotein receptor expressed on the liver.³)

We focused on the interaction between sialyl Lewis X (SLe^{X} , $Neu5Ac\alpha2\rightarrow3Gal\beta1\rightarrow4(Fuc\alpha1\rightarrow3)GlcNAc)$ and E-selectin. SLe^{X} is known to be a ligand of the cell adhesion molecule called E-selectin, which is expressed on the surface of endothelial cells during inflammation.⁴) As the interaction between SLe^{X} and E-selectin is essential for the initial stage of neutrophil infiltration into the inflammatory site, SLe^{X} and its derivatives, which block the interaction, should be useful as new anti-inflammatory agents.⁵)

Moreover, SLe^x and its derivatives may be considered to be effective homing devices for active-targeting DDS (<u>D</u>rug <u>D</u>elivery <u>S</u>ystem) to inflammatory lesions, since E-selectin is only expressed on such lesions. However, as carbohydrate recognition becomes understood, the affinity of carbohydrate for their protein has been shown to be relatively weak and carbohydrates to be generally sensitive to glycosidase *in vivo*. Furthermore, SLe^x should be rapidly filtered out at the glomerulus because of its high hydrophilicity and low molecular weight. One way to solve these problems would be to support SLe^x on a macromolecule? which would be stabilize the sugar moiety and multivalent interaction of SLe^x with E-selectin expressed at the inflammatory lesion. We have already reported that some polysaccharides with molecular size above 70KDa such as carboxymethylchitosan (CMCht)8) and carboxymethylpullulan (CMPul)9) are useful as carriers in passive-targeting DDS to tumors.9),10) The doxorubicin-CMPul conjugate gives dramatic enhancement of the therapeutic index of antitumor effects.9) This type of conjugate has high biocompatibility, is retained in blood circulation and accumulates in the tumor.



We report here the synthesis of SLe^{X} -polysaccharide (CMCht and CMPul) conjugates and evaluation of the possibility of using these conjugates as an active-targeting system for delivery to inflammatory lesions. We also synthesized $SLN(Neu5Ac\alpha2\rightarrow 3Gal\beta1\rightarrow 4GlcNAc)$ -polysaccharide conjugates as the negative control. SLN is a trisaccharide without the fucose moiety from SLe^{X} and has been reported to not support E-selectin-mediated adhesion. 11)

The synthesis of a SLe^{x} -CMCht conjugate (1) is described in Scheme 1. Glycosylation of 7^{12}) with 813) in presence of Me₂SSMe·OTf ¹⁴) in CH₂Cl₂ at 0°C (95%, α : β = 15:1), and subsequent regioselective opening of benzylidene ring using NaBH₃CN-HCl¹⁵), affords 9 (77%). However, there is a disadvantage with this ring opening reaction. NaBH₃CN is a hazardous chemical that precludes its safe use on a large scale. We investigated this reaction with various acids and found that Et₃SiH-TfOH¹⁶) is useful for this reaction. Using this system, compound 9 was cleanly obtained in 83% yield. Coupling of 9 with sialyl-galactose imidate (11)¹⁷) in the presence of BF₃·OEt₂ led to tetrasaccharide 12 (61%), which was hydrogenated with 10% Pd-C in MeOH to afford 13 (77%) and 14 (10%). Deacetylation of 13 followed by saponification of the methyl ester group gave 15 (93%). Similarly, 14 was converted to 5 (quant.).

Bromide (15) was reacted with CMCht⁸) (molecular weight: about 100kDa) in 0.5% NaHCO₃-H₂O for 160 hr at 60°C (*N*-alkylation) to afford a SLe^x-CMCht conjugate (1). SLe^x content of 1 was 33 wt%, and the degree of substitution (ds) of SLe^x was 0.17 per glucosamine residue of CMCht.¹⁸)

To synthesize the SLe^X-CMPul conjugate (2), treatment of 9 with LiN₃ afforded 10 (90%), which was coupled with imidate 11 (BF₃·OEt₂ / CH₂Cl₂) to give 16 (52%). Hydrogenation of 16 with 10% Pd-C in THF-HCl, gave amine 17 (89%).

Scheme 1

- a) 1) Me₂SSMe•OTf / CH₂Cl₂ / 0°C (95%, α : β =15:1) 2) Et₃SiH / TfOH / CH₂Cl₂ / MS4A / -78°C (87%)
- b) LiN₃ / DMF (90%) c) BF₃•OEt₂ / CH₂Cl₂ / MS4A / 0°C (12: 61%, 16: 52%)
- d) Pd-C / H₂ / MeOH / rt (77%) e) 1) NaOMe / MeOH / rt, 2) aq.NaOH / rt (15:93%, 5: quant.)
- f) Pd-C / H₂ / THF-HCI / rt (89%) g) 1) NaOMe / MeOH / rt, 2) aq.NaOH / rt,
- 3) Pd-C / H₂ / MeOH / pTsOH (93%) h) EtCO₂Su / MeOH / NMM (90%)

Condensation of amine 17 with CMPul⁹) (molecular weight: about 190kDa, ds of carboxymethyl group was 0.6 per glucose residue.) using 1-ethoxy-carbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ) in DMF-H₂O at 40°C, and subsequent deprotection with 1 N aq. NaOH, gave a SLe^x-CMPul conjugate (2), which was determined to contain 31 wt% SLe^x by the same method as 1.¹⁸) (Ds of SLe^x was 0.13 per glucose residue of CMPul.)

A SLN-CMCht conjugate (3) and a SLN-CMPul conjugate (4) were synthesized similarly. (3, ds = 0.17, 4, ds = 0.13, Scheme 3).

a)1) BnBr / BaO / Ba(OH) $_2$ *8 H_2 O / DMF / rt (85%) 2) Et $_3$ SiH / TfOH / CH $_2$ Cl $_2$ / MS4A / -78°C (87%) b) LiN $_3$ / DMF (90%) c) same procedure as **1** and **2**

The ability to actively target the inflammatory lesion *in vivo* was evaluated using the model of arachidonic acid-induced ear edema in mice. ¹⁹⁾ Drastic enhancement of accumulation in the targeted lesion was observed. AUC (Area Under the blood concentration-time Curve) 0-24 hr of macromolecules 1 and 2 at the inflammatory lesion was about 60-fold and 300-fold higher than that of the monovalent SLe^x derivative (6)²⁰), respectively. Less accumulation of CMCht conjugate 1 compared to 2 was due to the degradation of CMCht by lysozyme *in vivo*. Moreover, 1 showed 2-fold higher accumulation in the inflammatory lesion than 3, and 2 showed 2.5-fold higher accumulation than 4. We confirmed drug arrival at the E-selectin expressed at the inflammatory lesion by examining the inhibition of leukocyte rolling²¹)in the inflammatory model using IL-1 β activated rat mesentery venules. (*In vivo*) The multivalent SLe^x conjugate 2 was shown to have 6-fold higher inhibitory activity than the SLe^x analogue (5). (IC₅₀ value; 1: 0.57 μ M, 2: 0.40 μ M, 5: 2.63 μ M)

In summary, SLe^x-polysaccharide conjugates were shown to be a useful novel system for active-targeting DDS to inflammatory lesions. We are presently trying to introduce anti-inflammatory agents to SLe^x-polysaccharide conjugates and evaluate their increased therapeutic effects.

Acknowledgement

We thank Mr. Hideo Nogusa and Mr. Shuichi Sugawara for the gifts of CMCht and CMPul.

References and Notes

- 1) Present address: (a) Shionogi & Co., Ltd., Fukushima-ku, Osaka 553, Japan; (b) Eisai Co., Ltd., Tsukuba-shi, Ibaraki 300-26, Japan; (c) Tanabe Seiyaku Co., Ltd., Toda-shi, Saitama, Japan.
- (a) Imai, Y.; Lasky, L. A.; Rosen, S. D. Nature 1993, 361, 555.
 (b) Kelm, S.; Schauer, R.; Crocker, P. R. Glycoconjugate J. 1996, 13, 913.
 (c) Nakayama, J.; Fukuda, M. N.; Fredette, B.; Ranscht, B.; Fukuda, M. Proc. Natl. Acad. Sci. U.S.A. 1995, 92, 7031.
 (d) Perillo, N. L.; Pace, K. E.; Seihamer, J. J.; Baum, L. G. Nature 1995, 378, 736.

- (a) Ashwell, G.; Morell, A. Adv. Enzymol. 1974, 41, 99. (b) Spanjer, H. H.; Scherphof, G. L. Biochem. Biophys. Acta 1983, 734, 40. (c) Sasaki, A.; Murahashi, N.; Yamada, H.; Morikawa, A. Biol. Pharm. Bull. 1994, 17, 680.
- (a) Phillips, M. L.; Nudelman, E.; Gaeta, F. C. A.; Perez, M.; Singhal, A. K.; Hakomori, S.; Paulson, J. C. Science 1990, 250, 1130.
 (b) Walz, G.; Aruffo, A.; Kolanus, W.; Bevilacqua, M.; Seed, B. Science 1990, 250, 1132.
 (c) Springer, T. A.; Lasky, L. A. Nature 1991, 349, 196.
 (d) Bevilacqua, M.; Pober, J. S.; Mendrick, D. L.; Cotran, R. S.; Gimbrone, M. A., Jr. Proc. Natl. Acad. Sci. U.S.A. 1987, 84, 9238.
 (e) Bevilacqua, M.; Stengelin, S.; Gimbrone, M. A., Jr.; Seed, B. Science 1989, 243, 1160.
 (f) Lowe. J. B.; Stoolman, L. M.; Nair, R. P.; Larsen, R. D.; Berhend, T. L.; Marks, R. M. Cell 1990, 63, 475.
- 5) Uchiyama, T.; Vassilev, V. P.; Kajimoto, T.; Wong, W.; Huang, H.; Lin, C. -C.; Wong, C. -H. J. Am. Chem. Soc. 1995, 117, 5395.
- (a) Brenner, B. M.; Hostetter, T. H.; Humes, H. D. Am. J. Physiol. 1978, 234(6), F455.
 (b) Chang, R. L. S.; Ueki, I. F.; Troy, J. L.; Deen, W. N.; Brenner, B. M. Biophys. J. 1975, 15, 887.
- 7) (a) DeFrees, S. A.; Philips, L.; Guo, L.; Zalipsky, S. J. Am. Chem. Soc. 1996, 118, 6101. (b) Miyauchi, H.; Tanaka, M.; Koike, H.; Kawamura, N.; Hayashi, M. Bioorg. Med. Chem. Lett. 1997, 7, 985. (c) Thoma, G.; Magnani, L.; Ohrlein, R.; Ernst, B.; Schwarzenbach, F.; Duthaler, R. O. J. Am. Chem. Soc. 1997, 119, 7414.
- 8) Inoue, K.; Ito, T.; Okuno, S.; Aono, K. Patent WO/923480. The characters of CMCht show the following: degrees of substitution (ds) of *N*-actyl group: 0.47, ds of amino group: 0.45, ds of carboxymethyl group: 0.7.
- 9) Nogusa, H.; Yano, T.; Okuno, S.; Hamana, H.; Inoue, K. *Chem. Pharm. Bull.* **1995**, *43*, 1931. CMPul was prepared from Pullulan by carboxymethylation with chloroacetic acid and aq. NaOH.
- 10) Takakura, Y. Pharm. Res. 1987, 4, 293-300.
- 11) Tyrrell, D.; James, P.; Rao, N.; Foxall, C.; Abbas, S.; Dasgupta, F.; Nashed, M.; Hasegawa, A.; Kiso, M.; Asa, D.; Kidd, J.; Brandley, B. K. Proc. Natl. Acad. Sci. U.S.A. 1991, 88, 10372.
- 12) 7 was prepared from 2-methyl-4,5-(3,4,6-tri-*O*-acetyl-2-deoxy-α-D-glucopyrano)-2-oxazoline in 3 steps.[i) TMSOTf / CH₂Cl₂ / H-(OCH₂CH₂)6-Br (80%) ii) NaOMe / MeOH (quant.) iii) PhCH(OMe)₂/ CSA /DMF (65%)]
- 13) Yamazaki, F.; Sato, S.; Nukada, T.; Ito, Y.; Ogawa, T. Carbohydr. Res. 1990, 201, 31.
- 14) (a) Ravenscroft, M.; Roberts, R. M. G.; Tillett, J. G. J. Chem. Soc. Perkin Trans. II 1982, 1569. (b) Fügedi, P.; Garegg, P. J. Carbohydr. Res. 1986, 149, C9.
- 15) Garegg, J.; Hultberg, H.; Wallin, S. Carbohydr. Res. 1982, 108, 97.
- 16) Although the ring-opening reaction of benzylidene acetal using Et₃SiH-TFA has already been reported, our conditions using TfOH gave a much better yield. DeNinno, M. P.; Etienne, J. B.; Duplantier, K. C. *Tetrahedron Lett.* **1995**, *36*, 669.
- 17) Murase, T.; Kameyama, A.; Kartha, K. P. R.; Ishida, H.; Kiso, M.; Hasegawa, A. J. Carbohydrate Chemistry 1989, 8(2), 265.
- 18) SLe^X content was determined by the resorcinol-HCl method for quantitative analysis of sialic acid. a) Svennerholm, L. *Biochim. Biophys. Acta.* 1957, 24, 604. b) Miettinen, T.; Takki-Luukkainen, I. T. *Acta Chem. Scand.* 1959, 13, 856.
- 19) This evaluation was performed using radiolabelled 1-4. 1 and 3 were radiolabeled using *N*-succinimidyl [2,3-3H]-propionate. 2 and 4 were radiolabeled using [3H]-glycine and EEDQ. (Radioactivity: 1 and 3, 5 μCi / mg, 2 and 4, 15 μCi / mg)
- 20) Radiolabelled **6** was prepared from **18** using *N*-succinimidyl [2,3-³H]-propionate.
- 21) Olofsson, A. M.; Arfors, K.; Ramezani, L.; Wolitzky, B. A.; Butcher, E. C.; von Andrian, U. H. *Blood*, **1994**, 84, 2749. Leukocyte rolling in venules of the rat mesentery was induced by IL-1β. Experimental procedure are briefly shown in the following; Rat mesentery venules were treated by IL-1β, and after 2hr, test compounds were administered intravenously. After the interval of the 1hr, the number of the rolling leukocytes were measured during 1 min at rat mesentery venules with diameters of approximate 50 μm.